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COMPOSITION, NITRIFICATION AND FERTILIZING EFFECT OF ANAEROBICALLY FERMENTED SLURRY

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ABSTRACT

Under laboratory conditions, manure slurry from cattle (bulls), pig and chicken was fermented in 5 l digestors (batch system; 35°C; 5, 10, or 15 weeks).

Fermentation resulted in changes within the N or P fractions. With longer periods of fermentation, a decrease in organic N and P was observed. Decomposition of phytate P was significant only in chicken slurry.

Nitrification of slurry is primarily determined by the low C content after fermentation. In an incubation trial with ¹³N-labelled slurry-NH₄+ (soil: silty loam, pH 6.5; simulated top-soil temperatures during winter season) application of fermented slurry resulted in reducted biological N immobilization in the soil and consequently in a quick turnover of NH₄-N into NO₃-N.

In pot trials (I: Silty loam, pH 6.5; II: Sandy loam, pH 5.8, P20scal = 3 mg/100 g soil) fermented slurry gave partially higher yields when compared on the basis of equal amounts of NH4-N or P20s; in trial no.I these differences in yields disappeared with the subsequent crop because of a better long-term N supply from untreated slurry.

INTRODUCTION

During anaerobic fermentation of manure slurry only insignificant nutrient losses occur (Besson et al., 1982 a). By bacterial turnover a certain part of the organically bound nutrients is mineralized and an increased fertilizing effect of the fermented product can be expected (Koriath et al.,

These investigations were done to elucidate changes in N and P fractions during fermentation. In an incubation trial and in pot trials the nitrification rates and fertilizing effects were compared between anaerobically fermented and untreated slurry.

MATERIAL AND METHODS

1. Material

Chicken slurry (CS), bull slurry (BS) and pig slurry (PS) used in incubation and pot trials were fermented in 5 l digestors under laboratory conditions (batch system; 0, 5, 10, 15 weeks).

2. Methods

a) Determinations in slurry
 pH: in aqueous dilution with glass electrode (Orion Model 701);
 total C: wet ashing (Lichterfelder method) after Schlichting and Blume (1966).

Nitrogen:

Total nitrogen: after Kjeldahl;

Ammonium N: with ion-selective electrode (Orion Model 95-12); Hydrol.N: digestion (15 h) with 6 N HCl, filtration, Kjeldahl digestion of the filtrate;

Nonhydrol. N: Kjeldahl digestion of the residue.

HCl-soluble P: after John (1969); H₂0-soluble P: after Murphy and Riley (1962); Total P: wet ashing, determination after Gericke and Kurmies (1952);

column (Dowex^R-1, Cl⁻-form, 1×4 -200) after Harland and Oberleas Phytin-P: Extraction (1977); determination in the eluate after Latta and Eskin (1980); with 1 N HCl: separation on an anion exchange

b) Determination in soils

lation without (NH4+) or with addition of Devarda reagent (NO₃-) and after Kjeldahl digestion of the soil (total N) and titration with 0.005 M $\rm H_2SO_4$ (Vilsmeier and Medina, 1984); analysis by emission spectrometry in K2SO4 extract after NaOH distil-

3. Experimental outline

a) Incubation trial

capacity; bottles with 150 g soil (silty loam, pH 6.5, 30 ₽ slurry-15NH4-N, adjusted to 60% of the full water 2 mm fine) mixed in 500 ml polyethylene

Slurries: from bulls, fermented for 0 or 15 weeks.

Incubation: 0, 2, 5, 12, 18, 22 weeks. Temperature: adapted to top soil temperatures in the winter season:

b) Pot experiments

Experiment I: N fertilizing effect - slurry incorporated before seed:

Plants: green oat, ryegrass (2 cuts); Soil: silty loam, pH 6.5, 6.3 kg per pot;

Basic fertilizing: P, K optimal;

Fertilizer treatments:

control (without slurry)

3. 0.3 - 1.2 g N as NH₄NO₃.2. 0.3, 0.6, 0.9 g NH4-N as slurry

Experiment II: P fertilizing effect

Soil: sandy loam, pH 5.8, $P_2O_{scal} = 3 \text{ mg/}100 \text{ g}$, 6.3 kg per

Plants: green oat, ryegrass (8 cuts);

irregular intervals; Basic fertilizing: N, ~ optimal, according to demand =

Fertilizer treatments:

control (without slurry)

2. 0.4, 0.8 g P20s as freeze-dried slurry

0.4, 0.8 g P205 as dicalciumphosphate.

for an extended period to evaporate residual free ammonia.) (To exclude a N fertilizing effect, the freeze-dried slurry was aerated

RESULTS AND DISCUSSION

While pH pH value, dyry matter contents (dr.m.), N and P fractions (Table 1) values of all analyzed slurries except chicken slurry increase in

Table 1: Chemical composition of anaerobically fermented slurries

		cs*) 			PS.	1			8S *	Slurry, weeks*
	О		1	10 7			1	10 8			1 1
N	8.2	W	.9	7.3	7	•	0	8.0	•	ù	PH d
3.3	8	6.9	4.9	4.8	5.0	6.2	4.4	4.4	5.1	7.6	%
12.05	10.80	6.10	6.22	6.22	5.78	4.95	9.54	9.23	8.17	5.41	Total N fra
	0.79			0.71				0.72			N fractions (Total NH4-N/ N total N
0.03	0.12	0.23	0.21	0.18	0.18	0.32	0.14	0.20	0.24	0.27	(dr.m.) Hydr.N/
2.3	2.5	4.8	6.7	7.4	7.9	9.0	3.6	4.2	5.3	7.7	C /N
4.34	4.20	3.02	2.62	2.29	2.19	2.23	1.77	1.50	1.30	0.99	Total
0.25	0.16	0.24	0.13	0.55	0.45	0.31	0.16	0.20	0.31	0.49	fractions (dr.m.) H ₂ O-P/ HCl-P/P total P total P
0.86	0.83	0.73	0.73	0.71	0.68	0.77	0.81	0.71	0.73	0.55	(dr.m. HCl-P/ total P
69	98	390	12	16	14	19	14	17	26	36	Phytin p mg/100g

fermentation period

lower than untreated slurries. the course of fermentation, dry matter contents of fermented slurries are

crease in hydrolyzable N can be observed (chicken slurry). This N fraction slurry ammonium N comes up to nearly 90 % of total N. Accordingly, a decally bound nitrogen leads to a lower ratio of NH4-N/total N; of the soil (Amberger et al., 1982). is hardly plant available and almost completely enters the organic-N pool concentration effect in the fermented substrate. Mineralization of organi-Decomposition of organic matter and volatilization of biogas results in a in chicken

chicken slurry mainly due to a volatilization of ammonia (high pH value!). After Besson et al. (1982 b) these losses can add up to over 10 % depen-Balancing the analyzed N fractions reveals significant N losses only for ding on storage conditions.

and Mg phosphates (Fordham and Schwertmann, 1978). Decomposition of phytin more or less intensive P mineralization is observed (Gerritse, 1981). slurry). cause of high original concentrations. flects a mineralization of less soluble organic-P compounds (excluding pig The increase in the HCl-soluble fraction (up to 15 % more) of total P reis observed in all slurries, but is important only in chicken slurry bein H₂O-soluble P is caused by precipitation of hardly soluble Ca This is not surprising since even during normal slurry storage a

2. Nitrification of ammonia N from fermented slurry (Table 2)

on of the added slurry N and the remineralization of blocked N after a few pounds is applied. This is the reason for the decreased biological fixati-With fermented slurry a smaller amount of microbially available C com-Higher NOs concentrations in the soil extract of fermented slurry

of Rauhe et al. (1973) and Amberger et al. (1982). ciently reduced the formation of nitrate from slurry N in the experiments A strong microbial N fixation after straw manuring (C supply) also effi-

BS (bull slurry), PS (pig slurry), CS (chicken slurry)

Table 2: Nitrification of slurry-¹³NH₄-N at various incubation temperatures (simulation of winter season)

Bull slurry /	weeks	Temp.	NH*-N	NO FON	Residual N	Recovery rate	rate
0	2	4	54.8	9.8	35.4	100.0	
15	Ŋ	4	53.3	13.1	31.6	98.0	
0	u	0	32.4	40.5	29.5	102.4	
15	ហ	0	27.2	58.8	15.8	101.8	
0	12	0	. 21.3	47.1	30.3	98.7	
15	12	0	23.8	59.3	18.8	101.9	
0	18	7	11.8	62.2	26.2	100.2	
15	18	7	20.2	64.0	17.7	101.8	
0	22	11	0.5	76.9	22.0	99.4	
15	22	11	0.8	86.9	14.8	102.3	

* fermentation period

The rate of immobilization of 15 % or 22 % at the end of the experiment is relatively low compared to values of other authors (Peschke and Markgraf, 1982; Flowers and Arnold, 1983).

Nutrient effect of anaerobically treated cattle and pig slurry in pot trials

After incorporation fermented slurry has a somewhat better N fertilizing effect on the first cut (green oat), and untreated slurry has better effects on the subsequent crops (ryegrass, 2 cuts). Altogether, fermented slurry gives slightly inferior results (Fig. 1). This might be explained by a lower microbial immobilization of easily available ammonium N in the fermented slurry (viz. incubation trial) and a higher concentration of organic residual-N in the untreated slurry. To what extent this organic N is mineralized and plant available depends largely on ecological conditions in the soil and can hardly be predicted (Amberger, 1982).

The poor growth with pig slurry was caused by the very high application

The poor growth with pig slurry was caused by the very high application rates (due to the low ammonium concentrations in fresh matter) which resulted in structural damage probably followed by considerable N losses through denitrification. The good long-term N-supplying capacity of untreated slurry however can still be recognized.

In comparison, Koriath et al. (1985) reported significantly higher yields in one-year pot trials with maize after addition of anaerobically treated slurry, on the basis of equal amounts of total slurry N.

The obviously improved P fertilizing effect of fermented slurry (removal of P is generally increased with one exception: Fig. 2) is not surprising

ly bound slurry P during fermentation has already been pointed out. The use of freeze-dried NH₄*-free slurry did not completely exclude an additional residual effect of organic matter. However, since the experiment was done on a uniformly high N level, the influence caused by a masked N fertilizing effect is negligible.

because an influence on P forms or a possible mineralization of organical-

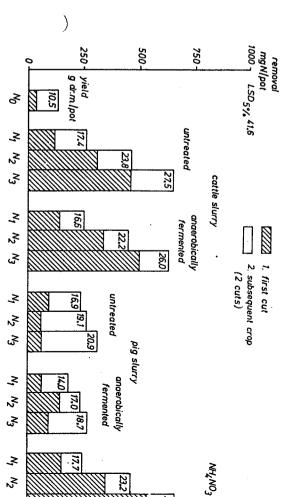


Fig.1: Effect of N from untreated and anaerobically fermented cattle and pig slurry

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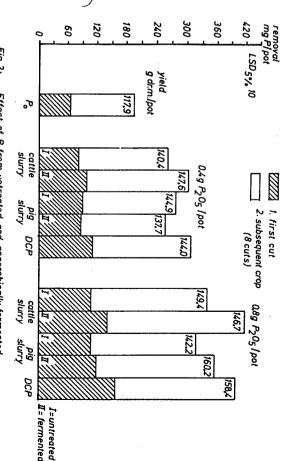


Fig.2: Effect of P from untreated and anaerobically fermented cattle and pig slurry

CONCLUSION

Anaerobic fermentation leads to mineralization of organic N and P compounds in slurry. As a consequence fermented slurry shows a long-term N-supplying capacity inferior to the untreated one. The low C/N ratio due to fermentation exerts an influence on nitrification. The reduced biological fixation of slurry N might favor possible leaching and translocation of nitrate into deeper soil layers. Anaerobically fermented slurry,

therefore, $% \left(1\right) =\left(1\right) +\left(1\right)$

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